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A *Drosophila* DEG/ENaC Subunit Functions Specifically in Gustatory Neurons Required for Male Courtship Behavior

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Detection of specific female pheromones stimulates courtship behavior in *Drosophila melanogaster* males, but the chemosensory molecules, cells, and mechanisms involved remain poorly understood. Here we show that *ppk25*, a DEG/ENaC ion channel subunit required for normal male response to females, is expressed at highest levels in a single sexually dimorphic gustatory neuron of most taste hairs on legs and wings, but not in neurons that detect courtship-inhibiting pheromones or food. Synaptic inactivation of *ppk25*-expressing neurons, or knockdown of *ppk25* expression in all gustatory neurons, significantly impairs male response to females, whereas gustatory expression of *ppk25* rescues the courtship behavior of *ppk25* mutant males. Remarkably, the only other detectable albeit significantly weaker expression of *ppk25* occurs in olfactory neurons implicated in modulation of courtship behavior. However, expression of *ppk25* in olfactory neurons is not required for male courtship under our experimental conditions. These data show that *ppk25* functions specifically in peripheral taste neurons involved in activation of courtship behavior, an unexpected function for this type of channel. Furthermore, our work identifies a small subset of gustatory neurons with an essential role in activation of male courtship behavior, most likely in response to female pheromones.

Introduction

In *Drosophila melanogaster*, initiation and modulation of the intricate courtship steps that precede mating involve complex communication between males and females (Villella and Hall, 2008; Dahanukar and Ray, 2011). While olfactory and gustatory receptors, sensory neurons, and even some of the central circuitry involved in detection and response to courtship-inhibiting pheromones have been identified (Kurtovic et al., 2007; Lacaille et al., 2007; van der Goes van Naters and Carlson, 2007; Datta et al., 2008; Laughlin et al., 2008; Miyamoto and Amrein, 2008; Yew et al., 2009; Koganezawa et al., 2010; Ronderos and Smith, 2010; Ruta et al., 2010; Wang et al., 2011), the cells and molecules involved in detecting pheromones that activate rather than inhibit courtship behavior are less understood, at least in part due to the involvement of several sensory modalities (Ejima and Griffith, 2008; Griffith and Ejima, 2009; Krstic et al., 2009). Both vision and hearing stimulate courtship initiation, but are not

essential, while pheromone detection plays a central role. Olfactory neurons expressing the olfactory receptor *Or47b* detect unknown fly odors and contribute to a male's ability to find a female and initiate courtship (van der Goes van Naters and Carlson, 2007; Root et al., 2008; Wang et al., 2011). However, loss of olfaction by itself has only a modest effect on overall levels of courtship (Stockinger et al., 2005; Krstic et al., 2009). In contrast, mutants lacking functional taste hairs display significant reductions in courtship, and males lacking both olfactory and gustatory inputs display almost no response to females (Boll and Noll, 2002; Krstic et al., 2009). In addition, since the only characterized pheromones that activate male courtship are long chain unsaturated cuticular hydrocarbons with very low volatility, their detection may require direct contact with gustatory organs (Ferveur et al., 1997; Ferveur, 2005; Siwicki et al., 2005; Wicker-Thomas, 2007). Two members of the *Gr* family of gustatory receptors are important for male response to females; however, the cells in which these two genes function and the sensory modality involved are either unknown or controversial (Bray and Amrein, 2003; Ejima and Griffith, 2008; Watanabe et al., 2011). Therefore, while contact perception of female pheromones is likely critical to activation of male courtship behavior, the gustatory neurons involved have not been identified.

We previously reported that mutations inactivating *ppk25*, a DEG/ENaC sodium channel subunit, impair activation of male courtship behavior in the presence of a female, but have no effect on behaviors unrelated to courtship, including walking, preening, and gustatory response to sugars and salts (Lin et al., 2005). Here we identify a small subset of gustatory neurons on the legs and wings that are specifically required for normal male courtship of females, and may therefore detect courtship-stimulating

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T.L. and V.V. contributed equally to this work.

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female pheromones. We further show that these gustatory neurons represent the site of *ppk25*'s function in activation of courtship behavior. These studies reveal that in addition to DEG/ENaCs that function in gustatory perception of sodium ions (Schiffman et al., 1983; Heck et al., 1984; Chandrashekar et al., 2010) or water molecules (Cameron et al., 2010; Chen et al., 2010); *ppk25* plays an unexpected role in gustatory activation of courtship behavior.

Materials and Methods

Generation of *ppk25-Gal4*, Western blots, immunostaining, and RNA-ligase-mediated rapid amplification of cDNA ends. The *ppk25-Gal4* construct was generated starting from a derivative of the pP[CaSpeR-4] (Qian et al., 1991) vector containing additional rare restriction enzyme cutting sites (our unpublished data). The insert contains four distinct fragments in the following order (Fig. 1B): (1) a 5 kb fragment with sequences upstream of the *ppk25* start codon and flanked by synthetic *AscI* and *PmeI* sites; (2) the yeast *Gal4* gene (Brand and Dormand, 1995) flanked by *PmeI* sites; (3) a 3832 bp *PmeI*-*NotI* fragment containing the entire *ppk25* coding region, inactivated by mutation of the ATG start codon to AAG, and ~2 kb of sequences downstream of the *ppk25* stop codon; and (4) a *NotI*-*BamHI* fragment of the pGATB vector containing the SV40 polyadenylation sequences (Brand and Dormand, 1995). Transgenic lines were generated using standard procedures (Rubin and Spradling, 1982). Immunostaining of antennae and brains was performed as described previously (Laisue et al., 1999) using rabbit anti-GFP (Invitrogen) at a dilution of 1:200, and anti-nc82 (Developmental Studies Hybridoma Bank, supported by the NICHD at the University of Iowa) at a dilution of 1:40. Western blots were performed with the same rabbit anti-GFP, and mouse anti- α -tubulin (Sigma-Aldrich), at 1:500 and 1:1000 dilutions, respectively. For RNA analysis, body fractions (200 male front legs or 100 heads, without antennae) were dissected by hand, the RNA extracted as published (Starostina et al., 2009), and RNA-ligase-mediated rapid amplification of cDNA ends (RLM-RACE) performed using a kit from Ambion.

Behavioral assays. Flies were raised at 25°C, 50% relative humidity in a 12 h:12 h light:dark cycle, except where indicated. For courtship assays, males were collected soon after eclosion and aged individually for 5–7 d before testing. Fifteen to twenty virgin wild-type females (either Canton-S or *w¹¹¹⁸*) were aged together for 5–7 d, then anesthetized and decapitated. After a short recovery, decapitated females that stood upright and preened were placed in the presence of a test male, and behavior was recorded with a digital Mini-DV camera for 10 min. Except where indicated, observation was performed under infrared light in Plexiglas chambers (7 mm diameter × 7 mm deep). Behaviors were scored blind and analyzed using the LIFESONG X software (version 0.8) (Vilella et al., 2005). The Courtship Index (CI) is the fraction of time the male spends performing any courtship behavior × 100 (Hall, 1978), while we defined a Total Behavioral Index (TBI) as the fraction of the same 10 min period during which the male courts, walks, or preens × 100. We find that the TBI provides a more objective measure of overall male activity than more specific parameters, e.g., measures of locomotor behavior, since reductions in courtship typically result in increased time spent performing behaviors unrelated to courtship but have little effect on the total behavior displayed. For CI and TBI, error bars indicating a 95% confidence interval were calculated as the SEM, and the Wilcoxon rank sum test was used to determine statistical significance. For the fraction of males initiating courtship, error bars indicate 95% confidence intervals for binomial distributions, and statistical significance was calculated using a χ^2 test for equality of distributions.

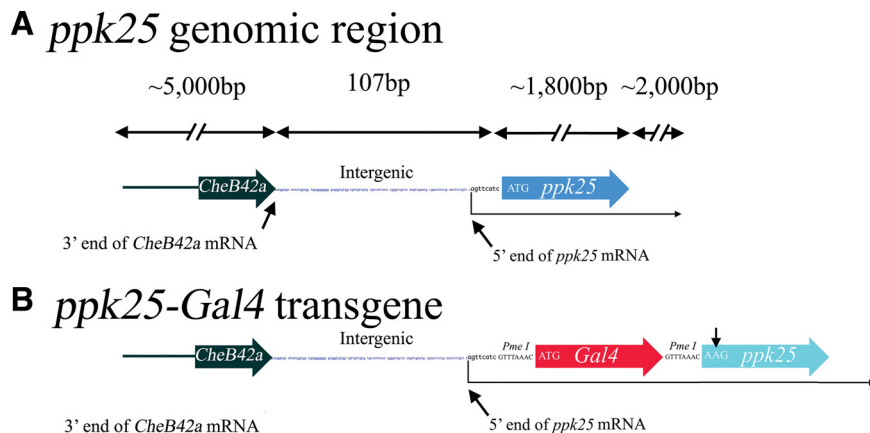


Figure 1. The *ppk25* gene region and construction of the *ppk25-Gal4* transgene. **A**, Map (not to scale) of the *ppk25* gene region and all genomic sequences included in the *ppk25-Gal4* transgene. Approximately 9 kb of sequences from the *ppk25* locus are included, including all sequences required for the functional rescue of courtship behavior in *ppk25* mutant males (Lin et al., 2005). This construct contains 4 kb of sequences upstream of the 5' end of the *CheB42a* mRNA (Xu et al., 2002) and all transcribed *CheB42a* sequences, and includes all regulatory sequences necessary for the functional expression of *CheB42a* (Park et al., 2006). The entire *ppk25* gene is also included, as well as 2 kb downstream of the *ppk25* open reading frame. Wide arrows indicate the *CheB42a* (black) and *ppk25* genes (blue). The 3' end of *CheB42a* mRNA (Xu et al., 2002) and the 5' end of the *ppk25* mRNA are indicated (see Results; Fig. 2A) (Ben-Shahar et al., 2007). **B**, Schematic representation of the *ppk25-Gal4* transgene. The wide red arrow represents the *Gal4* open reading frame (Brand and Dormand, 1995), inserted at the position of the *ppk25* ATG, 8 nt downstream of the 5' end of the *ppk25* mRNA, and a light blue arrow indicates the *ppk25* coding region inactivated by elimination of its start codon.

Gustatory response to sucrose was assayed by measuring the proboscis extension reflex (PER), as described previously (Heimbeck et al., 2001). Briefly, 5- to 7-d-old males were starved for 24 h in vials without food but containing water-soaked cotton wool. After 24 h, males were anesthetized with CO₂, attached to glass slides on their backs using myristic acid (Acros Organics), and allowed to recover for 2 h in saturating humidity. Each live fly was then given water until it no longer exhibited a PER when touched on its forelegs with a drop of water, at which point water was replaced with a 100 mM sucrose solution and the response tested five times with a delay of 30 s between tests to avoid habituation (Heimbeck et al., 2001). The PER index represents the number of times the male extended its proboscis in response to sucrose divided by 5.

***Drosophila* stocks.** Most fly lines used were obtained from the Bloomington Stock Center. Two RNAi lines were obtained from the Vienna *Drosophila* RNAi Center (stock numbers v101808 and v41512, targeting *ppk25* and CG13895, respectively), and one RNAi line from the Transgenic RNAi Project at Harvard Medical School targeting *ppk25* (stock number JF02434). The *Poxn-Gal4*, *fru-LexA*, and *lexAop-FRT-tdTomato::nls*; UAS-*stinger* lines were gifts from David Mellert.

Results

A *ppk25-Gal4* transgene reflects expression of the endogenous *ppk25* gene

To define the expression and function of *ppk25* using the UAS/*Gal4* system (Elliott and Brand, 2008), we generated a transgenic construct in which the yeast transcriptional activator *Gal4* is placed under control of *ppk25* regulatory regions. For many *Drosophila* genes, 3 kb of upstream sequences are sufficient to direct reporter gene expression in the pattern of the endogenous gene (Pfeiffer et al., 2008). In the case of *ppk25*, however, only 107 nt separate the 3' end of the *CheB42a* mRNA from the ATG initiator codon for the Ppk25 protein (Lin et al., 2005; Ben-Shahar et al., 2007) (Fig. 1A). Given this unusually short intergenic region, sequences required for correct regulation of *ppk25* expression may occur upstream of the *CheB42a* gene or within it, or within the *ppk25* gene itself, perhaps in an intron or in its 3' untranslated region. To best reproduce the context of the original *ppk25* gene, we therefore inserted the *Gal4* open reading frame immediately after the *ppk25* start codon, within a large genomic fragment that

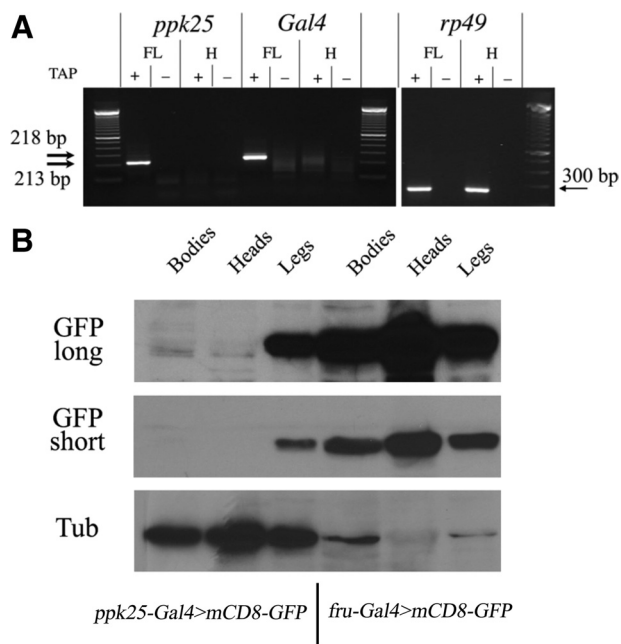


Figure 2. Both *ppk25-Gal4* and the endogenous *ppk25* gene are expressed in legs but not heads. **A**, Transcripts of the *ppk25-Gal4* transgene and of the endogenous *ppk25* gene initiate at the same nucleotide, and are found in front legs but not in heads. RNA extracted from front legs (FL) or heads without antennae (H) was analyzed by amplification of cDNAs using RLM-RACE, which only amplifies cDNAs produced by reverse transcription of 5'-capped RNAs (Schaefer, 1995). + and – indicate whether the RNA sample was treated with tobacco acid pyrophosphatase (TAP) to generate a 5' monophosphate required for ligation to the RNA adaptor. Specific antisense primers were used for *ppk25*, *Gal4*, and *rp49*. The unique bands of 218 and 213 bp observed in cDNAs from front leg and resulting from amplification of the *ppk25* and *Gal4* transcripts, respectively, were sequenced, confirming the reported start site for the *ppk25* mRNA (Ben-Shahar et al., 2007), and an identical start site for the *ppk25-Gal4* transgene (Fig. 1). In contrast, the ubiquitous *rp49* mRNA yields a predicted band of 300 bp that is detected equally by RLM-RACE using cDNA from front legs and heads. **B**, Expression of GFP under control of *ppk25-Gal4* is detectable in legs but not heads or bodies. Extracts were made from bodies (without legs, wings, or heads), heads (without antennae), or legs of males expressing mCD8-GFP under control of either *ppk25-Gal4* or *fru-Gal4* (Stockinger et al., 2005) as indicated, and analyzed by immunoblotting with anti-GFP. The blots were then stripped and reprobed with anti-tubulin antibody (Tub), showing that for males containing the *ppk25-Gal4* driver, more extract from heads than legs was loaded on the gel. For those males, extracts from heads and bodies show no detectable GFP band above the background of nonspecific bands. GFP short and GFP long refer to different exposures of the same Western blot.

contains all *ppk25* regulatory sequences required to rescue *ppk25* function in a deletion mutant (Lin et al., 2005). In this construct, the *Gal4* gene is preceded by 5 kb of sequences normally found upstream of *ppk25*, including the entire *CheB42a* gene and all sequences required for its regulation (Xu et al., 2002; Park et al., 2006), and is followed by the entire *ppk25* coding region, inactivated by a point mutation in the initiator codon, and ~2 kb of noncoding downstream sequences (Fig. 1B).

To test whether expression of *ppk25-Gal4* accurately reflects that of the endogenous *ppk25* gene, we compared the sites of transcription initiation and the tissue distribution of the two mRNAs using RLM-RACE, a method that only amplifies cDNAs resulting from reverse transcription of 5'-capped mRNAs (Schaefer, 1995). Specific primers allow amplification of the 5' end, either of the endogenous *ppk25* transcripts, or of transcripts from the *ppk25-Gal4* transgene, yielding a single detectable RACE product in each case (Fig. 2A). Direct sequencing of the PCR product resulting from the *ppk25* mRNA shows that the initiation site is just 8 nt upstream of the ATG (Figs. 1A, 2A), confirming a previous report (Ben-Shahar et al., 2007). Furthermore, sequenc-

ing of the PCR product resulting from the *ppk25-Gal4* mRNA (Fig. 2A) indicates that transcription of this transgene initiates at the same nucleotide as the endogenous gene (Fig. 1B). In addition, RACE products for both *ppk25* and *ppk25-Gal4* can be detected in the legs but not in a head fraction from which the third antennal segment was removed, mirroring the distribution of full-length *ppk25* mRNA detected by Northern blot (Lin et al., 2005), whereas expression of the ubiquitously expressed *rp49* is detected equally in both fractions. These results show that the *ppk25* and *ppk25-Gal4* mRNAs initiate at the same nucleotide and are expressed in the same body parts, suggesting that expression of the *ppk25-Gal4* driver reflects that of the endogenous *ppk25* gene.

To further test the specificity of *ppk25-Gal4* expression, and in particular its lack of expression in the brain, we used Western blots to compare levels of GFP in extracts of different body parts (Fig. 2B). Here also we find that *ppk25-Gal4*-driven GFP expression is readily detectable in the legs, but not in the heads (without antennae), or in the bodies of flies. In contrast, use of *fru-Gal4*, a driver predominantly expressed in the brain (Manoli et al., 2005; Stockinger et al., 2005), results in high levels of GFP in head extracts (Fig. 2B). Together, these data suggest that the *ppk25-Gal4* transgene and the endogenous *ppk25* gene are regulated in similar ways, making *ppk25-Gal4* a powerful tool to study the function of *ppk25* and the cells in which it functions.

Expression of *ppk25-Gal4* is restricted to small characteristic subsets of gustatory and olfactory neurons, some of which have been implicated in male courtship behavior

Drosophila gustatory neurons are located within taste hairs found mostly on the mouthparts, legs, and wings (Fig. 3A) (Shanbhag et al., 2001; Vosshall and Stocker, 2007). Under control of *ppk25-Gal4*, mCD8-GFP expression is observed in most taste hairs of the front legs (Fig. 3B,C), other legs (data not shown), and wings (Fig. 3D) of both males and females, but in no other part of the body (data not shown). Targeted expression of tau-GFP, a marker of neuronal processes in *Drosophila* (Murray et al., 1998) (Fig. 3C,C'), allows visualization of a long apical dendrite through the entire hair shaft (Fig. 3C', green arrow), identifying these cells as gustatory neurons (Shanbhag et al., 2001). Interestingly, double-staining with anti-CheB42a antibody (Park et al., 2006) shows that expression of the two neighboring genes does not overlap despite the very short intergenic region (Fig. 1A); instead, *ppk25* expression occurs in gustatory neurons, whereas, as shown previously (Park et al., 2006), *CheB42a* is expressed in the surrounding non-neuronal sheath cells (Fig. 3E,F). The expression of *CheB42a* and *ppk25* in distinct cell types and the presence of a 5' cap modification on the *ppk25* mRNA as indicated by its amplification by RLM-RACE (Fig. 2A), together with the absence of a detectable precursor RNA containing both *CheB42a* and *ppk25* sequences (Park et al., 2006; Ben-Shahar et al., 2007), indicate that the adjacent *CheB42a* and *ppk25* genes are transcribed independently rather than resulting from the post-transcriptional processing of a common polycistronic transcript as previously suggested (Ben-Shahar et al., 2007).

Expression of *ppk25-Gal4* defines a small subset of gustatory neurons with distinguishing characteristics. Most taste hairs on the legs contain four gustatory neurons (Shanbhag et al., 2001). Two of these taste neurons express *fruitless* (*fru*), the master regulator of neuronal sexual determination required for most male-specific behaviors (Siwicki and Kravitz, 2009), and send characteristic sexually dimorphic projections onto the prothoracic ganglia (Mellert et al., 2010). Expression of *ppk25-Gal4* is

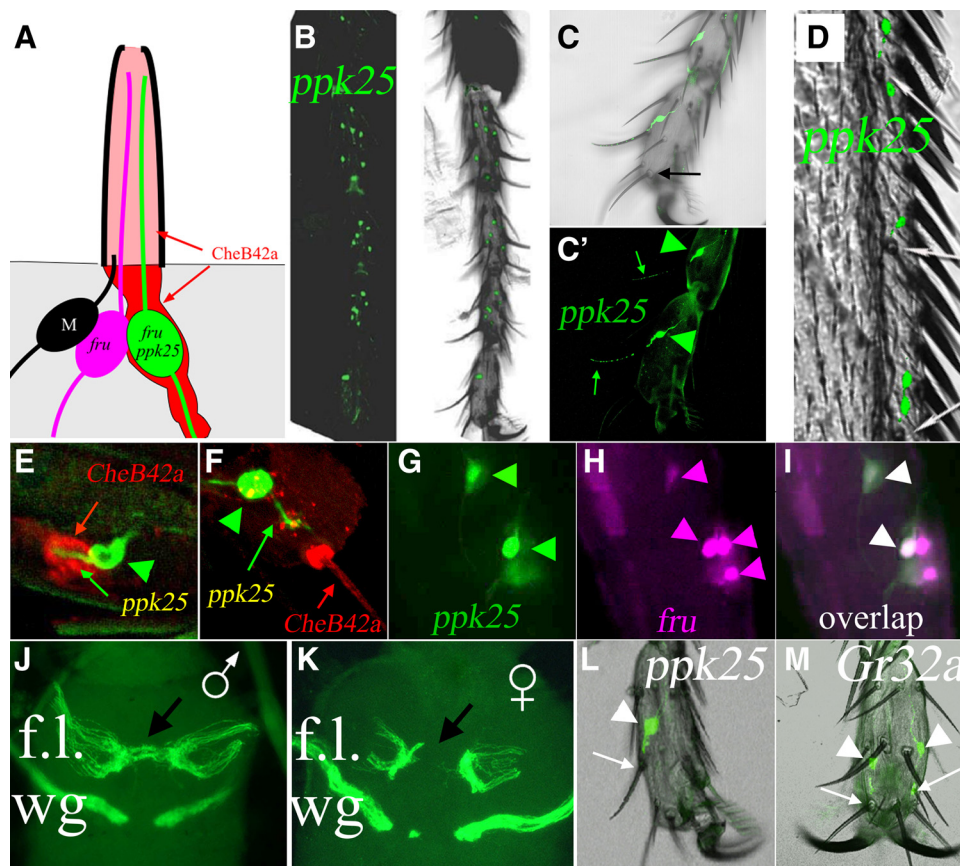


Figure 3. *ppk25-Gal4* is expressed in a small subset of gustatory neurons implicated in sexual behavior. **A**, Schematic drawing of a gustatory hair of the leg. The dendrites of two to four gustatory neurons (only the two *fru*-expressing neurons are shown) extend through the hair lumen to the terminal pore where they can interact with chemicals in the environment (Shanbhag et al., 2001). One neuron expresses both *ppk25* and *fru* (green), while another expresses *fru* but not *ppk25* (magenta; see Results). A single mechanosensory neuron (M, black) in each hair has much shorter, stout dendrites that terminate at the base of the hair (Shanbhag et al., 2001). The CheB42a protein is expressed in non-neuronal cells (red) that surround the *ppk25*-expressing neuron and its dendrites and is secreted into the lumen of the taste hair (Park et al., 2006; Starostina et al., 2009) (see Results). **B**, Expression of membrane-tethered mCD8-GFP under control of the *ppk25-Gal4* driver is present in most taste hairs of all legs (male foreleg shown). **C**, Higher magnification shows that expression is confined to only one gustatory neuron per taste hair. **C** and **C'** show superimposed fluorescent and bright light images, or fluorescence only, respectively. In this experiment, *ppk25-Gal4* drives expression of tau-GFP, a reporter protein that localizes to neurites (Murray et al., 1998). In each of two GFP-expressing cells visible in this field, a dendrite extends through the whole length of the hair shaft (**C'**, green arrow), indicating that expression occurs in gustatory neurons. Of six taste hairs in the terminal leg segment (Meunier et al., 2000, 2003), only two hairs in the proximal part of the segment house a mCD8-GFP-labeled gustatory neuron (only one of which is seen in this optical section). There is no reporter gene expression in taste hairs closer to the leg tip which contains food-sensing neurons (one such hair in focus in **C** is indicated by a black arrow) (Meunier et al., 2000, 2003). **D**, In the wing also, *ppk25-Gal4* drives expression of mCD8-GFP in a single gustatory neuron in each taste hair. **E, F**, CheB42a, a male-specific gene immediately upstream of *ppk25* (Fig. 1A) is expressed in non-neuronal support cells (Park et al., 2006; Starostina et al., 2009), in the same taste hairs as *ppk25-Gal4*-expressing gustatory neurons. **E**, CheB42a-expressing non-neuronal sheath cells labeled with anti-CheB42a antibody (red) surround a gustatory neuron that expresses mCD8-GFP under control of *ppk25-Gal4* (green, cell body indicated by arrowheads; dendrite by arrows). **F**, Optical section through the shaft of a taste hair showing secreted CheB42a in the lumen of a taste hair penetrated by the dendrite of a *ppk25-Gal4*-expressing neuron. **G–I**, Taste neurons expressing nuclear GFP under control of *ppk25-Gal4* (**G**, in green) also express tdTomato::nls under control of *fru-LexA* (Mellert et al., 2010) (**H, I**, tdTomato::nls in magenta, overlap of tdTomato::nls and GFP in white). **J, K**, The taste neurons expressing mCD8-GFP under control of *ppk25-Gal4* in the front legs send sexually dimorphic projections to the first thoracic ganglia. Gustatory neurons expressing *ppk25-Gal4* in the front legs (f.l.) make projections that cross the midline in males (**J**, arrow), but not females (**K**), as reported for *fru*-expressing gustatory neurons (Possidente and Murphey, 1989; Boll and Noll, 2002; Mellert et al., 2010). In contrast, the projections from *ppk25-Gal4*-expressing neurons on the wings (wg) and from the second and third pairs of legs (data not shown) do not cross the midline in flies of either sex. **L, M**, *ppk25* and *Gr32a*, gustatory receptors that inhibit courtship behavior (Miyamoto and Amrein, 2008; Koganezawa et al., 2010; Wang et al., 2011), are expressed in nonoverlapping patterns. Observations of the fifth tarsal segment show that dendrites of *ppk25-Gal4*-expressing gustatory neurons (only one of two *ppk25-Gal4*-expressing neurons is visible in this side view) penetrate the shaft of taste hairs in the center of the tarsal segment (**L**, cell body indicated by arrowhead, arrow points to base of the corresponding taste hair), whereas dendrites of gustatory neurons that express *Gr32a-Gal4* enter taste hairs that are closer to the leg tip (**M**, cell bodies indicated by arrowhead, arrows point to bases of the corresponding taste hairs).

restricted to a single gustatory neuron per taste hair (Fig. 3C,E–G). Identification of the *ppk25*-expressing cell as one of the *fru*-expressing neurons is demonstrated by double-labeling (Fig. 3G–I) and confirmed by the sexual dimorphism of its projections onto the prothoracic ganglia (Fig. 3J,K). Intriguingly, *ppk25-Gal4* is not expressed in the tips of legs, the site of several taste hairs that respond to general food components such as sugars and bitter compounds (Fig. 3C, black arrow) (Meunier et al., 2000, 2003), but is expressed in most other leg taste hairs that do not respond to food, some of which have been implicated in detection of pheromones that either stimulate or inhibit courtship

behavior (Bray and Amrein, 2003; Park et al., 2006; Miyamoto and Amrein, 2008; Koganezawa et al., 2010; Lee et al., 2010; Wang et al., 2011; Watanabe et al., 2011). The specificity of *ppk25-Gal4* expression is further highlighted by its absence from the many taste hairs present in other parts of the adult body (Shanbhag et al., 2001; Vosshall and Stocker, 2007; Montell, 2009). In particular, *ppk25-Gal4* is not expressed in the numerous gustatory hairs found on the mouthparts (data not shown), most of which have been shown to respond to food stimuli such as sugars, salts, and bitter compounds. Finally, comparison of the expression of *ppk25* to that of *Gr32a*, a gustatory receptor that inhibits male–

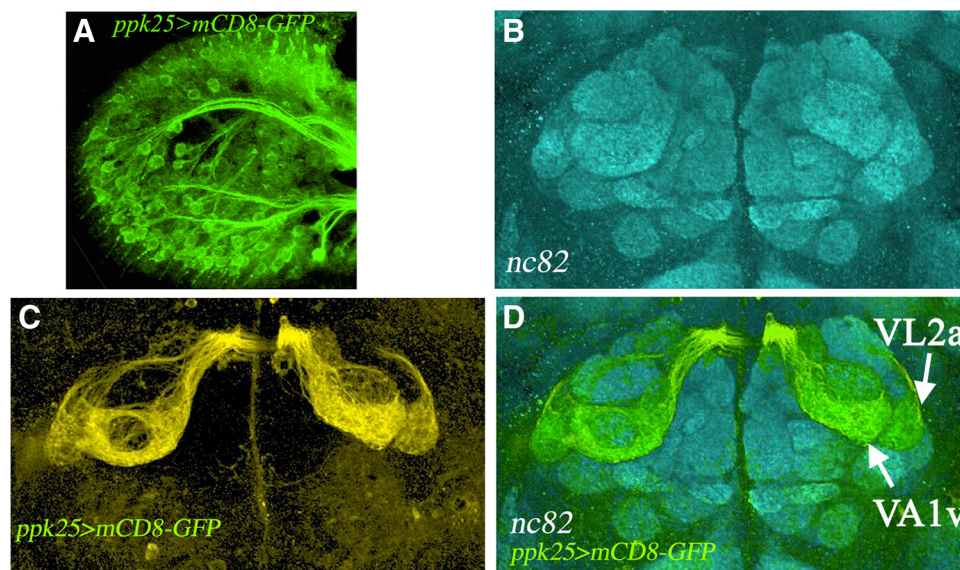


Figure 4. *ppk25-Gal4* is expressed in two subsets of olfactory neurons that stimulate male courtship. **A**, Immunostaining reveals *mCD8-GFP* expression in olfactory neurons within the third antennal segment of *ppk25-Gal4;UAS-mCD8-GFP* males. **B**, Whole-brain immunostaining of males of the same genotype with neuronal-specific *nc82* decorates the glomeruli of the antennal lobe. **C**, Staining with anti-GFP reveals the axonal projections of *ppk25-Gal4*-expressing neurons. **D**, A superimposition of the two immunostaining patterns shows that the antennal neurons that express *ppk25-Gal4* project to two glomeruli that are easily identified by their position on the antennal lobe and characteristic shapes: VA1v, also called VA1Im, and VL2a (Couto et al., 2005; Fishilevich and Vosshall, 2005). Both VA1v and VL2a are sexually dimorphic and express *fru* (Kondoh et al., 2003; Stockinger et al., 2005). VA1v receives projections from olfactory neurons that respond to female cuticular extracts and express *Or47b*, an olfactory receptor for fly odors (van der Goes van Naters and Carlson, 2007; Root et al., 2008; Wang et al., 2011), while VL2a is innervated by olfactory neurons that express *IR84a* and stimulate courtship in response to aromatic food odors (Grosjean et al., 2011).

male courtship (Miyamoto and Amrein, 2008; Koganezawa et al., 2010) in response to 7-tricosene (Wang et al., 2011), suggests that the two genes are expressed in different subsets of leg gustatory neurons (Figs. 3*L,M*), consistent with the absence of *fruitless* expression in *Gr32a*-expressing neurons (Miyamoto and Amrein, 2008; Koganezawa et al., 2010). While *ppk25-Gal4* drives *mCD8-GFP* expression in all five tarsal segments on the front legs (Fig. 3*C* and not shown), *Gr32a-Gal4*-driven expression is observed only in the fourth and fifth segments ((Miyamoto and Amrein, 2008; Koganezawa et al., 2010) (data not shown). On the fifth segment, *ppk25-Gal4* labels two neurons that innervate hairs in the middle of the segment, whereas *Gr32a-Gal4* is expressed in two neurons that innervate hairs close to the tip of that segment (3 *M*; Fig. 3*L*, arrowheads indicate the neuronal cell bodies; arrows point to the base of the hair penetrated by the dendrite). In the fourth tarsal segment, counts of cell bodies that express *mCD8-GFP* under control of *ppk25-Gal4*, *Gr32a-Gal4*, or in the presence of both transgenes consistently revealed 7, 3, and 10 taste neurons, respectively, ($n > 10$ for each genotype; data not shown), suggesting that here also, expression of the two drivers does not overlap. Finally, we do not observe *ppk25-Gal4*-directed expression of *mCD8-GFP* in chemosensory neurons of the ovipositor (Stocker, 1994), and the absence of *mCD8-GFP*-labeled axonal projections to the abdominal ganglia (data not shown) suggests that *ppk25-Gal4* is not expressed in the *fru*-positive internal sensory neurons of the female reproductive tract (Häsemeyer et al., 2009; Yang et al., 2009).

To determine whether *ppk25-Gal4* directs lower levels of *mCD8-GFP* expression in other tissues, we used immunocytochemistry. Remarkably, the only additional reporter expression detected with this more sensitive method occurs in two subsets of antennal olfactory neurons, both of which have been implicated in olfactory stimulation of male courtship behavior (Fig. 4*A*). Forty-three distinct types of olfactory neurons have been described based on the expression of specific olfactory receptor (*Or*)

genes, each projecting to a single identifiable pair of bilaterally symmetrical glomeruli in the antennal lobe (Couto et al., 2005; Fishilevich and Vosshall, 2005). *ppk25-Gal4*-expressing olfactory neurons project only to two pairs of glomeruli, VA1v and VL2a, identified by their shape, size, and position within the antennal lobe (Fig. 4*B–D*). Both VA1v and VL2a are sexually dimorphic and express *fruitless* (Kondoh et al., 2003; Couto et al., 2005; Fishilevich and Vosshall, 2005; Manoli et al., 2005; Stockinger et al., 2005). Furthermore, both VA1v and VL2a have been implicated in stimulating male courtship behavior, the former through detection of fly odors (van der Goes van Naters and Carlson, 2007; Root et al., 2008; Wang et al., 2011), and the latter in response to aromatic food chemicals (Grosjean et al., 2011). In summary, expression of *ppk25-Gal4* is restricted to small subsets of gustatory and olfactory neurons. For both gustatory neurons of the legs and olfactory neurons of the antennae, *ppk25-Gal4* expression occurs in a subset of *fru*-expressing neurons and includes neurons with either known or suspected roles in stimulation of male courtship behavior.

A small subset of chemosensory neurons is required for male response to females

To test whether the chemosensory neurons defined by expression of *ppk25-Gal4* are important for male courtship behavior in response to females, we used targeted expression of *shi^{ts}*, a temperature-sensitive, dominant allele of *Dynamin* that blocks synaptic transmission at restrictive temperatures (Kitamoto, 2001) (Fig. 5). When kept at the permissive temperature of 18°C, males expressing *shi^{ts}* under control of *ppk25-Gal4* and control males expressing GFP in the same pattern display similar levels of courtship as measured by the CI (the fraction of a 10 min observation period spent performing any of the steps involved in the courtship sequence $\times 100$) (Hall, 1978). However, incubation for 1 h at the nonpermissive temperature of 30°C results in a 10-fold lower CI for males expressing *shi^{ts}* relative to controls. In con-

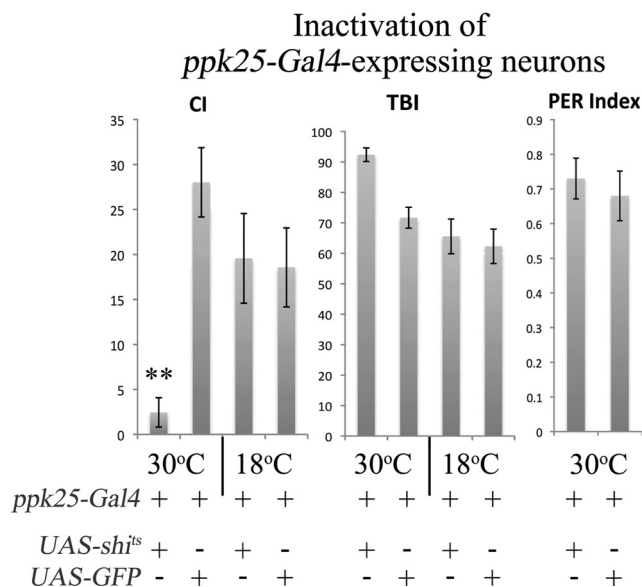


Figure 5. Inactivation of *ppk25*-expressing neurons specifically impairs male response to virgin females. Males expressing the temperature-sensitive dominant *Dynamin* allele *shi^{ts}* (Kitamoto, 2001) under control of *ppk25-Gal4*, were raised at the permissive temperature (18°C) and, where indicated, incubated for 1 h at the nonpermissive temperature (30°C) just before testing their behavior at the same temperature. Response to a Canton-S virgin female was recorded under white light for a 10 min interval. Initiation is the fraction of males to initiate courtship within the 10 min observation period. Calculation of the CI, the TBI, the PER, and the corresponding SEs and statistical tests are described in Materials and Methods. ** $p < 0.01$. For both genotypes, $N = 28$ and $N = 22$ at 30°C and 18°C, respectively.

trast, *shi^{ts}*-mediated inactivation of *ppk25-Gal4*-expressing neurons does not affect the TBI (fraction of time the test male spends performing any of the most common behaviors: courtship, walking, or preening, multiplied by 100), or gustatory response to sucrose as measured by the PER (Fig. 5, right) (Heimbeck et al., 2001). These results indicate that the peripheral chemosensory neurons that express *ppk25-Gal4* play an important and specific role in stimulating male courtship behavior in response to females.

ppk25 function in a small subset of gustatory neurons is required and sufficient for normal male courtship behavior in response to females

The similarity of the phenotypes caused by *ppk25-Gal4*-targeted expression of *shi^{ts}* and by mutations in *ppk25* (Lin et al., 2005) suggests that *ppk25* functions in the peripheral chemosensory neurons in which *ppk25-Gal4* is expressed. Indeed, *ppk25-Gal4*-directed expression of wild-type *ppk25*, but not of a mutant truncated *ppk25* (*ppk25^Δ*), is sufficient to restore a normal CI to *ppk25* mutant males (Fig. 6A), but does not affect other behaviors (Fig. 6B), consistent with a specific function for *ppk25* in courtship behavior (Lin et al., 2005). In contrast, expression of *ppk25* in olfactory neurons known to respond to a small number of food odors using the *Or22a-Gal4* driver (Hallem et al., 2004) has no effect on male courtship (Fig. 6A). These results suggest that the endogenous *ppk25* gene functions in the peripheral chemosensory neurons that express *ppk25-Gal4* and raise the question of whether this function occurs in gustatory neurons, in olfactory neurons, or in both types of sensory neurons. Interestingly, while *ppk25-Gal4* is expressed in olfactory neurons that project to the VA1v glomeruli which have been implicated in response to fly odors (van der Goes van Naters and Carlson, 2007; Root et al., 2008; Wang et al., 2011), targeted expression of *ppk25* in neurons

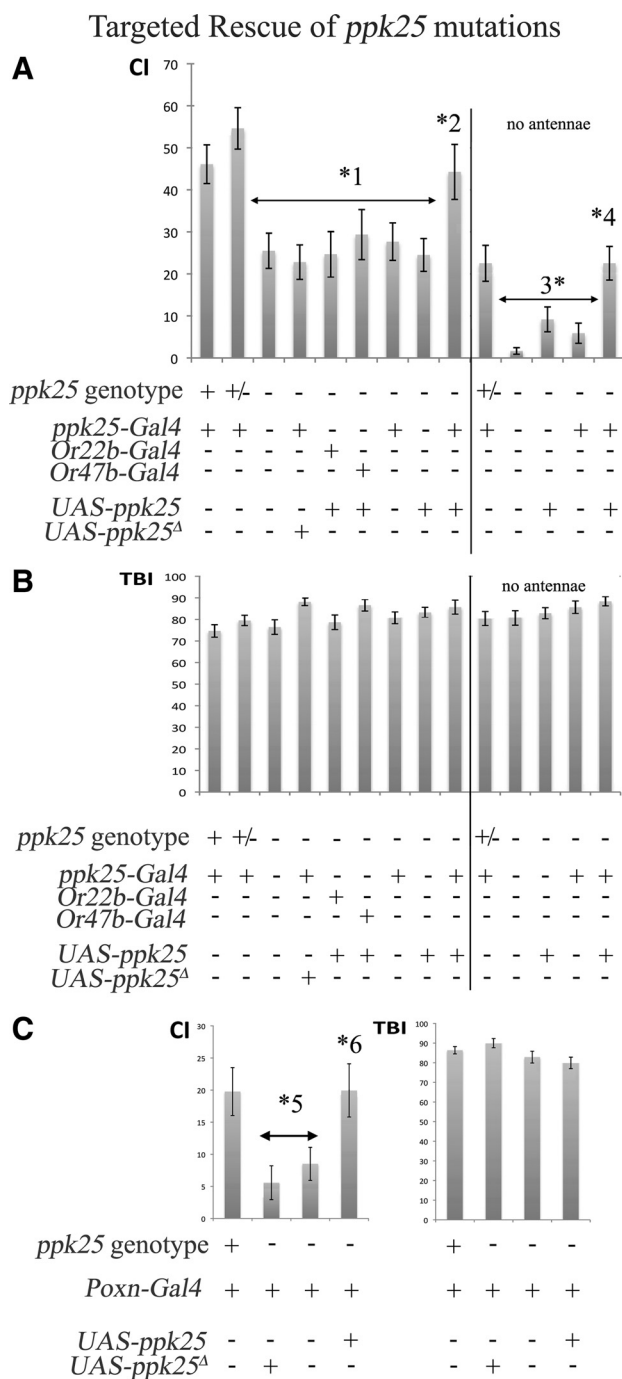


Figure 6. Targeted expression of *ppk25* in gustatory neurons rescues the courtship behavior of *ppk25* mutant males. Courtship behavior was measured for the males containing the transgenes indicated, and of the following genotypes: control males that are wild-type for *ppk25*, +; males heterozygous for a deletion spanning *ppk25*, +/−; males heterozygous for two different deletions spanning the *ppk25* gene, −. **A**, targeted expression of *ppk25* using *ppk25-Gal4* rescues male courtship. Right panel, labeled as “no antennae,” shows experiments in which the third antennal segment was removed bilaterally. In all cases, * indicates a statistically significant difference, $p < 0.05$. Statistically significant differences are as follows: *1, between control males carrying one or two copies of the wild-type *ppk25* gene and *ppk25* mutants with various combinations of transgenes; and *2, between homozygous *ppk25* mutants expressing *ppk25* under control of *ppk25-Gal4* and homozygous *ppk25* mutants carrying other combinations of transgenes. *3, *4 refer to the corresponding differences among antennaeless males. **B**, Males of all genotypes have a similar TBI. **C**, The *ppk25* mutant phenotype is rescued by targeted expression of *ppk25* in all gustatory neurons using the *Poxn-Gal4* driver (Boll and Noll, 2002). Statistically significant differences between control males and *ppk25* mutant males with various transgenes, and between *ppk25* mutants with various transgenes and males expressing *ppk25* under control of *Poxn-Gal4*, are indicated by *5 and *6, respectively. Here also, males of all genotypes have similar TBIs.

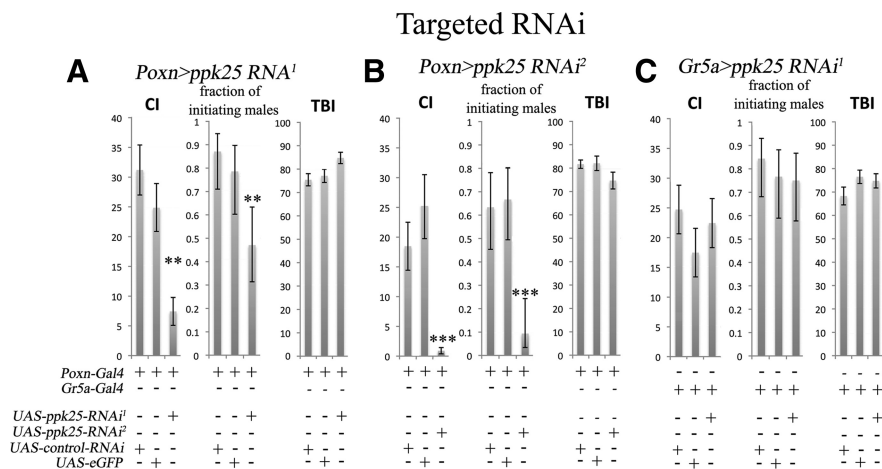


Figure 7. Targeted RNAi knockdown of *ppk25* in gustatory neurons blocks male courtship behavior. **A**, Gustatory expression of *ppk25 RNAi*¹ using the *Poxn-Gal4* driver (Boll and Noll, 2002) reduces both CI and the fraction of initiating males. Control males expressed either eGFP, or a control RNAi against *CG13895*, a gene with no known involvement in mating behavior (Benchabane et al., 2011), in the same pattern. **B**, Gustatory expression of a second RNAi transgene, *ppk25 RNAi*², using the *Poxn-Gal4* driver reduces both CI and the fraction of initiating males. This experiment was performed at 27°C because preliminary tests at 25°C resulted in effects that were not statistically significant at $p < 0.05$ (data not shown). **C**, As in **A**, except for the use of *Gr5a-Gal4* to drive expression in sucrose-sensing neurons of the labelum and legs (Chyb et al., 2003). Expression of *ppk25 RNAi*¹ in those neurons does not result in any significant effect on male courtship.

projecting to the same glomeruli using the *Or47b-Gal4* driver does not rescue the courtship of *ppk25* mutant males. To evaluate more directly the importance of the antennal expression of *ppk25*, we tested whether removing the main olfactory organ, the third antennal segment (Masse et al., 2009), affected *ppk25-Gal4*-directed rescue of *ppk25* mutants. Consistent with other reports, loss of olfaction reduces male response to females by approximately ~2-fold (Stockinger et al., 2005; Krstic et al., 2009) (Fig. 6A). More important, courtship of such antennaless males is further reduced in mutants lacking the *ppk25* gene and targeted expression of *ppk25* using *ppk25-Gal4* rescues their response to females to the level displayed by antennaless wild-type males, whereas the presence of either parental transgene alone has no significant effect (Fig. 6A). In contrast, neither removal of the antennae nor expression of functional *ppk25* affects the total behavioral output of males (Fig. 6B). These data suggest that in the absence of antennae at least, expression of *ppk25* in the small subset of *ppk25-Gal4*-expressing gustatory neurons on the legs and wings is sufficient for normal male courtship.

To further test whether *ppk25* functions in gustatory neurons, we asked whether targeted expression of *ppk25* using *Poxn-Gal4*, a driver expressed specifically and at high levels in all gustatory neurons (Boll and Noll, 2002), was sufficient to rescue the behavioral phenotype of *ppk25* mutant males (Fig. 6A). In preliminary experiments, we were surprised to find that in *ppk25* heterozygotes, the *Poxn-Gal4* driver itself inhibits male courtship behavior by ~2-fold (Fig. 6, compare A and C and data not shown), perhaps as a result of high levels of the Gal4 protein in gustatory neurons, as reported for other subsets of gustatory neurons (Lacaille et al., 2007; Yang et al., 2008), and other types of neurons (Rezaval et al., 2007). Nevertheless, comparison of a series of males that all carry the *Poxn-Gal4* driver shows that inactivation of the *ppk25* gene results in a significant further reduction of male response to females, and that *Poxn-Gal4*-directed expression of *ppk25*, but not of a truncated *ppk25*^Δ, rescues male behavior to the level found in the presence of the endogenous *ppk25* (Fig. 6C). Here also, males of all genotypes have similar TBIs (Fig. 6C). These experiments further support the conclusion that gustatory expression of *ppk25* is sufficient for normal male courtship.

In complementary experiments, we tested the effect on male courtship of knocking down the *ppk25* mRNA specifically in gustatory neurons using two different *UAS-ppk25 RNAi* transgenes (Dietzl et al., 2007; Ni et al., 2009) under control of *Poxn-Gal4*. Males expressing either *ppk25 RNAi* under control of *Poxn-Gal4* display severely reduced levels of overall courtship but normal levels of total behavior compared with control males expressing an unrelated RNAi or a GFP reporter in the same neurons (Fig. 7A,B). In contrast, expression of *ppk25 RNAi* in sugar-sensing gustatory neurons using the *Gr5a-Gal4* driver (Chyb et al., 2003) does not affect courtship behavior (Fig. 7C) or response to sucrose as measured by PER (data not shown). Furthermore, the effect of gustatory-specific knockdown of *ppk25* is not restricted to late steps in courtship behavior, as the fraction of males that initiate courtship is also severely reduced relative to controls (Fig. 7; $p < 0.01$ and $p < 0.001$ for *ppk25-RNAi*¹ and *ppk25-RNAi*², respectively). These results indicate that *ppk25* function in gustatory neurons is required for normal courtship, starting from the earliest steps in this behavioral sequence.

In summary, our data reveal that under our experimental conditions *ppk25* function in a small and specific subset of gustatory neurons on the legs and wings is both necessary and sufficient for normal activation of male courtship behavior in response to females.

Discussion

Identification of a subset of gustatory neurons required for activation of male courtship of females

Activation of the courtship behavior of *Drosophila melanogaster* males involves detection of specific female pheromones, including several well characterized hydrocarbons (Antony et al., 1985; Ferveur et al., 1997; Ferveur, 2005; Siwicki et al., 2005; Billeter et al., 2009). While the sense of smell plays a role in courtship activation (Villella and Hall, 2008), gustatory detection appears to be more important (Fig. 7) (Boll and Noll, 2002; Krstic et al., 2009). However, before this work, the gustatory neurons involved in detecting courtship-activating pheromones remained uncertain. Here we identify a small subset of gustatory neurons on the legs and wings of males that have an essential function in the activa-

tion of courtship behavior. Use of a *ppk25-Gal4* transgene to specifically inhibit synaptic function in small subsets of gustatory and olfactory neurons dramatically reduces male courtship of females, while expression of *ppk25* in the same neurons rescues the defective courtship of *ppk25* mutant males. Additional experiments further localize this function to the gustatory neurons on the legs and wings where *ppk25-Gal4* is expressed. First, the *ppk25* mutant phenotype is rescued by gustatory-specific expression of *ppk25* by two different methods: (1) using *ppk25-Gal4* to drive *ppk25* expression in males lacking their antennae, the site of the only detectable expression of *ppk25-Gal4* outside of the gustatory system, and (2) using the gustatory-specific *Poxn-Gal4* driver. Second, gustatory-specific knockdown of *ppk25* function using two distinct *ppk25* RNAs under control of *Poxn-Gal4* reduces the levels of male courtship displayed by factors of 7 and 20. Together, these results indicate that expression of *ppk25* in the small subsets of gustatory neurons defined by expression of *ppk25-Gal4* on the legs and wings of males are both required and under laboratory conditions, at least, are sufficient for normal activation of male courtship behavior.

These findings have implications not only for the gustatory function of *ppk25*, but also for its possible involvement in olfaction. The very low levels of courtship displayed by males after gustatory-specific RNAi-mediated knockdown of *ppk25* suggest that expression of *ppk25* in olfactory neurons is not sufficient to initiate courtship. Furthermore, since olfaction allows detection of volatile chemicals at a distance from their source, it has been generally assumed that olfactory detection is solely responsible for the effects of pheromones on the initiation of male courtship, while gustatory perception of pheromones becomes possible only after direct contact of the gustatory hairs of the male with the female body. The severe defects displayed by males carrying different *ppk25* alleles in both early and late events in courtship therefore led us to speculate that *ppk25* may function in both chemical senses (Lin et al., 2005). However, we find that specific knockdown of *ppk25* mRNA in taste neurons significantly reduces the fraction of males that initiate any courtship behavior (Fig. 7A,B). These data confirm a previous report that gustatory perception plays a key role not only in the maintenance, but also in the initiation of courtship behavior (Krstic et al., 2009). Gustatory stimulation may occur as the male and female touch by chance before any courtship behavior has been displayed, or alternatively as the male comes in direct contact with female pheromones deposited on the surface of the observation chamber in a mechanism analogous to the contact detection of low volatility trail pheromones in some ants and caterpillars (Roessingh et al., 1988; Jackson et al., 2006). These data indicate that *ppk25* functions in gustatory neurons that have an important role, not only in the late steps of male courtship, but also in the initiation of this complex behavior.

It is nonetheless remarkable that the only detectable expression of *ppk25-Gal4* outside of the gustatory system, albeit at a significantly lower level, occurs in two subsets of sexually dimorphic olfactory neurons, both of which express *fruitless* and stimulate courtship behavior either in direct response to female odors (van der Goes van Naters and Carlson, 2007; Wang et al., 2011), or through detection of aromatic fruit compounds that enhance the attractiveness of females (Grosjean et al., 2011). *ppk25* may therefore have a role in olfactory modulation of courtship that would only become apparent under conditions that enhance the importance of olfactory inputs (Ejima and Griffith, 2008; Root et al., 2008; Grosjean et al., 2011).

What are the ligand(s) detected by the gustatory neurons that express *ppk25-Gal4*? Synaptic inactivation of those neurons does not affect gustatory response to sugar (Fig. 5), or result in increased male–male courtship as seen in males deficient in detection of courtship-inhibiting pheromones (data not shown) (Kurtovic et al., 2007; Miyamoto and Amrein, 2008). In addition, expression of *ppk25-Gal4* does not occur on the proboscis or leg tips where many specialized food-sensing hairs are located. Finally, expression of *ppk25-Gal4* in the taste hairs of the legs only occurs in one of the two *fru*-positive neurons present, and does not overlap with that of *Gr32a*, a taste receptor that inhibits male courtship behavior (Miyamoto and Amrein, 2008) through detection of 7-tricosene (Wang et al., 2011). Together, these data suggest that the gustatory neurons defined by expression of *ppk25-Gal4* have a specialized role in detecting one or more ligands that activate courtship behavior. The relevant ligands remain to be identified, and it will be of particular interest to test whether they include one or more of the known low-volatility female pheromones that activate male courtship (Antony et al., 1985; Ferveur et al., 1997; Ferveur, 2005; Siwicki et al., 2005; Billeter et al., 2009).

A DEG/ENaC channel with a specific role in gustatory activation of courtship behavior

What is the molecular mechanism that underlies *ppk25*'s role in gustatory activation of male courtship? *ppk25* may have a direct role in gustatory detection of a courtship-stimulating molecule, either as part of a receptor complex, or further downstream in the signaling cascade. While such mechanisms deviate from the well known G-protein-coupled processes involved in vertebrate chemical senses (Pifferi et al., 2010), recent findings suggest that in insects, both olfaction and taste function through ion channels that are directly gated by environmental ligands and belong to multiple structurally distinct families (Silbering and Benton, 2010). In particular, another *Drosophila* DEG/ENaC subunit, *ppk28*, mediates gustatory response to water and its ectopic expression confers osmotic sensitivity to nonresponsive neurons, suggesting that gating of *ppk28* must be controlled by osmotic pressure (Cameron et al., 2010; Chen et al., 2010). In vertebrates, ENaC expressed in taste cells functions as a salt receptor, depolarization resulting directly from the specific translocation of extracellular sodium (Schiffman et al., 1983; Heck et al., 1984; Chandrasekar et al., 2010), and similar mechanisms may be involved for two *Drosophila* DEG/ENaCs implicated in salt response, *ppk11* and *ppk19* (Liu et al., 2003). Finally, ligand-gated DEG/ENaCs are also found outside of the gustatory system, e.g., protons and other ligands trigger opening of the acid-sensing channel (Yu et al., 2010), and FRMF-amide gates the gastropod FRMF-amide receptor (Poët et al., 2001). Gating of *ppk25* may therefore be controlled by courtship-activating molecules, either directly or in a complex with pheromone-binding proteins in a mechanism related to olfactory detection of *cis*-vaccenyl acetate, an inhibitory volatile pheromone (Laughlin et al., 2008). An alternative possibility is that *ppk25* may play a less direct role in modulating the response of courtship-activating gustatory neurons. By analogy to a *Caenorhabditis elegans* DEG/ENaC channel that mediates learning through presynaptic facilitation of dopamine release (Voglits and Tavernarakis, 2008), *ppk25* may exert its effect on neuronal function in the axon or at the synaptic terminal rather than in sensory dendrites. This work opens the way to the functional characterization of gustatory activation of male courtship, and in particular to the dissection of the molecular role

of *ppk25* and other DEG/ENaC channel subunits in gustatory detection of pheromones and other organic molecules.

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